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## Nucleosides, Nucleotides and Nucleic Acids

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## Synthesis of 2-Deoxy- $\beta$ -D-ribose 1-Phosphate, NMR Comparison and Its Enzymatic Activity for Structural Elucidation of Synthetic $\alpha$ -Isomer

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### ABSTRACT

2-Deoxy- $\beta$ -D-ribose 1-phosphate (**1**) was synthesized in a stereoselective manner and isolated with no detectable contamination by its  $\alpha$ -isomer (**4**). Explicit configuration of **4** was first determined by NMR comparison with **1** judging from NOE results and their coupling constants. Natural purine nucleoside phosphorylase (PNPase) did not recognize **1** and gave no products such as  $\alpha$ - or  $\beta$ -deoxynucleosides.

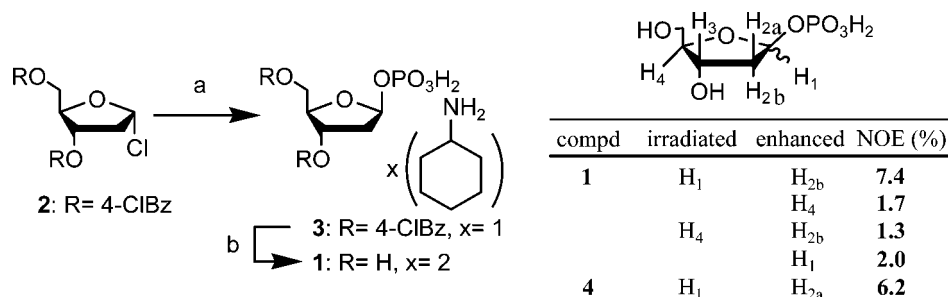
*Key Words:* Chemo-enzymatic; 2-Deoxyribose 1-phosphate.

### INTRODUCTION

A first stereoselective synthesis of 2-deoxy- $\alpha$ -D-ribose 1-phosphate (**4**)<sup>[1]</sup> has been established in our laboratory. Since **4** is a substrate for an enzymatic conversion into various 2'-deoxynucleosides (dNus), the result enabled us the development of

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**Figure 1.** Stereoselective synthesis of 2-deoxy- $\beta$ -D-ribose 1-phosphate (**1**), and NOE results of **1** its  $\alpha$ -isomer (**4**).<sup>[1]</sup> (a)  $o$ -H<sub>3</sub>PO<sub>4</sub>, Oct<sub>3</sub>N, MEK then  $c$ -C<sub>6</sub>H<sub>11</sub>NH<sub>2</sub>, 84%; (b)  $c$ -C<sub>6</sub>H<sub>11</sub>NH<sub>2</sub>, MeOH, 87%.

scalable processes for dNus.<sup>[2]</sup> Even though  $\alpha$ -selectivity of the synthetic reaction for **4** was significantly high, effects of the residual  $\beta$ -isomer (**1**) against the enzyme had been obscure for us. Therefore, an enzymatic reaction using **1** has been required for a direct evaluation to eliminate subliminal concern with the possible contamination by the  $\alpha$ -isomer of dNus. Additionally, since the absolute configuration of **4** has remained ambiguous,<sup>[3]</sup> it should be confirmed by spectroscopic analyses by comparing **4** with **1** (Fig. 1).

## RESULTS AND DISCUSSION

Nucleophilic substitution of chlorosugar (**2**) is rapid and undergoes inversion. MacDonald's method<sup>[3]</sup> was modified by using tri(*n*-octyl)amine (Oct<sub>3</sub>N) to increase the solubility of the corresponding H<sub>3</sub>PO<sub>4</sub> salt that effectively facilitated the substitution ( $\beta$ : $\alpha$ =90:10). Formation of cyclohexylamine ( $c$ -C<sub>6</sub>H<sub>11</sub>NH<sub>2</sub>) salt, followed by recrystallization from MeOH-acetone gave **3** in 84% yield with no detectable  $\alpha$ -isomer on HPLC assay. Finally, deprotection by  $c$ -C<sub>6</sub>H<sub>11</sub>NH<sub>2</sub> in MeOH gave **1** in 87% yield. It rotated  $\alpha_D^{25} = -22.2^\circ$  ( $c$  3, H<sub>2</sub>O) (lit.<sup>[3]</sup>  $\alpha_D^{20} = -15.8^\circ$  ( $c$  1.2, H<sub>2</sub>O)). NMR experiments of **1** and **4**,<sup>[1]</sup> such as Nuclear Overhauser Effect (NOE) and <sup>1</sup>H-<sup>1</sup>H spin decoupling supported the absolute configuration proposed by MacDonald that was based on the analogous property with  $\alpha$ - and  $\beta$ -D-ribose. In enzymatic reaction using adenine, **1** was not recognizable by natural PNPase and gave neither  $\alpha$ - nor  $\beta$ -dNus.

In summary, we showed the synthesis of **1** with no contamination by **4**. The result allowed us to confirm the absolute configuration of **1** and **4**, and to certify that **1** was not recognizable by natural PNPase.

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